



MAGLUMI® PIGF (CLIA)

INTENDED USE

The kit is an *in vitro* chemiluminescence immunoassay for the quantitative determination of placenta growth factor (PIGF) in human serum using the MAGLUMI series Fully-auto chemiluminescence immunoassay analyzer and Biolumi series Integrated System. It can be combined with other tests and clinical information to assist in the diagnosis of pre-eclampsia.

SUMMARY

Vascular endothelial growth factor (VEGF) represents a growth factor with important pro-angiogenic activity, having a mitogenic and an anti-apoptotic effect on endothelial cells, increasing the vascular permeability, promoting cell migration, etc. The VEGF family is composed of several members: VEGF-A, VEGF-B, VEGF-C, VEGF-D, VEGF-E, VEGF-F, PIGF, and recently, to this family has been added endocrine gland-derived vascular endothelial growth factor (EG-VEGF)¹. PIGF was first isolated and purified from human placental cDNA library in 1991. It is a protein containing 149 amino acids². PIGF is a secreted dimeric glycoprotein that shares significant homology with VEGF. PIGF is a potent angiogenic growth factor capable of inducing proliferation, migration, and activation of endothelial cells. Abundant expression of PIGF is restricted to the placenta, with the primary site of synthesis within the placenta being trophoblast³. At gestation of 29 to 32 weeks, PIGF reaches the maximum value, and then decreases⁴.

Preeclampsia is a common hypertensive disorder specific to pregnancy occurring in 3-5% of all pregnant women who are diagnosed by new onset hypertension and proteinuria occurring after 20 weeks of gestation⁵. The clinical onset of preeclampsia is often insidious and asymptomatic, but may include headache, visual disturbances, epigastric pain, weight gain, and edema of the hands and face⁶. At the same time, the blood coagulation function of preeclampsia is abnormal, mainly manifested by increased fibrin formation, activation of the fibrinolytic system, platelet activation and decreased platelet count⁷. Preeclampsia is the most common reason for iatrogenic preterm delivery⁸. Preeclampsia is a major cause of maternal and perinatal mortality and morbidity worldwide and it is generated by shallow invasion of the extravillous trophoblast followed by an incomplete remodeling of the maternal vascular structures which leads to uteroplacental insufficiency and intrauterine growth restriction, which in turn can influence placental angiogenesis and development⁹. In preeclampsia, PIGF concentrations began to decrease 9-11 weeks before the appearance of hypertension and proteinuria, with considerable diminution during the 5 weeks before the onset of disease. Therefore, in early second trimester, serum PIGF concentrations in preeclamptic pregnancies would be significantly lower than those of normotensive pregnancies⁷. Increased PIGF is a new independent predictor of cardiovascular morbidity and mortality in type 1 diabetic patients with diabetic nephropathy¹⁰. And PIGF is associated with plaque inflammation and formation of microvessels, important processes leading to plaque instability and symptomatic carotid disease¹¹.

TEST PRINCIPLE

Sandwich chemiluminescence immunoassay.

The sample, buffer, ABEI labeled with anti-PIGF monoclonal antibody and magnetic microbeads coated with another anti-PIGF monoclonal antibody are mixed thoroughly and incubated, reacting to form sandwich complexes. After precipitation in a magnetic field, the supernatant is decanted and then a wash cycle is performed. Subsequently, the Starter 1+2 are added to initiate a chemiluminescence reaction. The light signal is measured by a photomultiplier as relative light units (RLUs), which is proportional to the concentration of PIGF present in the sample.

REAGENTS

Kit Contents

Component	Description	100 tests/kit	50 tests/kit	30 tests/kit
Magnetic Microbeads	Magnetic microbeads coated with anti-PIGF monoclonal antibody (~8.00 µg/mL) in PBS buffer, NaNa ₃ (<0.1%).	2.5 mL	1.5 mL	1.0 mL
Calibrator Low	A low concentration of PIGF antigen in Tris-HCl buffer, NaNa ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Calibrator High	A high concentration of PIGF antigen in Tris-HCl buffer, NaNa ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Buffer	Tris-HCl buffer, NaNa ₃ (<0.1%).	6.5 mL	4.0 mL	3.0 mL
ABEI Label	ABEI labeled with anti-PIGF monoclonal antibody (~125 ng/mL) in Tris-HCl buffer, NaNa ₃ (<0.1%).	17.5 mL	9.5 mL	6.3 mL
Control 1	A low concentration of PIGF antigen (200 pg/mL) in Tris-HCl buffer, NaNa ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Control 2	A high concentration of PIGF antigen (1200 pg/mL) in Tris-HCl buffer, NaNa ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
All reagents are provided ready-to-use.				

Warnings and Precautions

- For *in vitro* diagnostic use.
- For professional use only.
- Exercise the normal precautions required for handling all laboratory reagents.
- Personal protective measures should be taken to prevent any part of the human body from contacting samples, reagents, and controls, and should comply with local operating requirements for the assay.
- A skillful technique and strict adherence to the package insert are necessary to obtain reliable results.
- Do not use kit beyond the expiration date indicated on the label.
- Do not interchange reagent components from different reagents or lots.
- Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).
- All waste associated with biological samples, biological reagents and disposable materials used for the assay should be considered potentially infectious and should be disposed of in accordance with local guidelines.
- This product contains sodium azide. Sodium azide may react with lead or copper plumbing to form highly explosive metal azides. Immediately after disposal, flush with a large volume of water to prevent azide build-up. For additional information, see Safety Data Sheets available for professional user on request.

Note: If any serious incident has occurred in relation to the device, please report to Shenzhen New Industries Biomedical Engineering Co., Ltd. (Snibe) or our authorized representative and the competent authority of the Member State in which you are established.

Reagent Handling

- To avoid contamination, wear clean gloves when operating with a reagent kit and sample. When handling reagent kit, replace the gloves that have been in contact with samples, since introduction of samples will result in unreliable results.
- Do not use kit in malfunction conditions; e.g., the kit leaking at the sealing film or elsewhere, obviously turbid or precipitation is found in reagents (except for Magnetic Microbeads) or control value is out of the specified range repeatedly. When kit in malfunction conditions, please contact Snibe or our authorized distributor.
- To avoid evaporation of the liquid in the opened reagent kits in refrigerator, it is recommended that the opened reagent kits to be sealed with reagent seals contained within the packaging. The reagent seals are single use, and if more seals are needed, please contact Snibe or our authorized distributor.
- Over time, residual liquids may dry on the septum surface. These are typically dried salts and have no effect on assay efficacy.
- Use always the same analyzer for an opened reagent integral.
- For magnetic microbeads mixing instructions, refer to the Preparation of the Reagent section of this package insert.
- For further information about the reagent handing during system operation, please refer to Analyzer Operating Instructions.

Storage and Stability

- Do not freeze the integral reagents.
- Store the reagent kit upright to ensure complete availability of the magnetic microbeads.
- Protect from direct sunlight.



0123



130212010M: 100 tests/kit
130612010M: 50 tests/kit
130712010M: 30 tests/kit

Stability of the Reagents	
Unopened at 2-8°C	until the stated expiration date
Opened at 2-8°C	6 weeks
On-board	4 weeks

Stability of Controls	
Unopened at 2-8°C	until the stated expiration date
Opened at 10-30°C	6 hours
Opened at 2-8°C	6 weeks
Frozen at -20°C	3 months
Frozen and thawed cycles	no more than 3 times

SPECIMEN COLLECTION AND PREPARATION

Specimen Types

Only the specimens listed below were tested and found acceptable.

Specimen Types	Collection Tubes
Serum	Tubes without additive/accessory, or tubes containing clot activator or clot activator with gel.

- The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. Follow tube manufacturers' instructions carefully when using collection tubes.

Specimen Conditions

- Do not use heat-inactivated samples or grossly hemolyzed/hyperlipidaemia specimens and specimens with obvious microbial contamination.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some serum specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the serum specimen is centrifuged before a complete clotting, the presence of fibrin may cause erroneous results.
- Samples must be free of fibrin and other particulate matter.
- To prevent cross contamination, use of disposable pipettes or pipette tips are recommended.

Preparation for Analysis

- Inspect all specimens for foam. Remove foam with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.
- Frozen specimens must be completely thawed before mixing. Mix thawed specimens thoroughly by low speed vortexing or by gently inverting. Visually inspect the specimens. If layering or stratification is observed, mix until specimens are visibly homogeneous. If specimens are not mixed thoroughly, inconsistent results may be obtained.
- Specimens should be free of fibrin, red blood cells, or other particulate matter. Such specimens may give reliable results and must be centrifuged prior to testing. Transfer clarified specimen to a sample cup or secondary tube for testing. For centrifuged specimens with a lipid layer, transfer only the clarified specimen and not the lipemic material.
- The sample volume required for a single determination of this assay is 50 µL.

Specimen Storage

Specimens removed from the separator, red blood cells or clot may be stored up to 6 hours at 10-30°C or 48 hours at 2-8°C, or 6 months frozen at -20°C. Frozen specimens subjected to up to 3 freeze/thaw cycles have been evaluated.

Specimen Shipping

- Package and label specimens in compliance with applicable local regulations covering the transport of clinical specimens and infectious substances.
- Do not exceed the storage limitations listed above.

Specimen Dilution

- Further dilution is not necessary due to the broad measuring range.

PROCEDURE

Materials Provided

PIGF (CLIA) assay, control barcode labels.

Materials Required (But Not Provided)

- General laboratory equipment.
- Fully-auto chemiluminescence immunoassay analyzer Maglumi 600, Maglumi 800, Maglumi 1000, Maglumi 2000, Maglumi 2000 Plus, Maglumi 4000, Maglumi 4000 Plus, MAGLUMI X3, MAGLUMI X6, MAGLUMI X8 or Integrated System Biolumi 8000 and Biolumi CX8.
- Additional accessories of test required for the above analyzers include Reaction Module, Starter 1+2, Wash Concentrate, Light Check, Tip, and Reaction Cup. Specific accessories and accessories' specification for each model refer to corresponding Analyzer Operating Instructions.
- Please use accessories specified by Snibe to ensure the reliability of the test results.

Assay Procedure

Preparation of the Reagent

- Take the reagent kit out of the box and visually inspect the integral vials for leaking at the sealing film or elsewhere. If there is no leakage, please tear off the sealing film carefully.
- Open the reagent area door; hold the reagent handle to get the RFID label close to the RFID reader (for about 2s); the buzzer will beep; one beep sound indicates successful sensing.
- Keeping the reagent straight insert to the bottom along the blank reagent track.
- Observe whether the reagent information is displayed successfully in the software interface, otherwise repeat the above two steps.
- Resuspension of the magnetic microbeads takes place automatically when the kit is loaded successfully, ensuring the magnetic microbeads are totally resuspended homogenous prior to use.

Assay Calibration

- Select the assay to be calibrated and execute calibration operation in reagent area interface. For specific information on ordering calibrations, refer to the calibration section of Analyzer Operating Instructions.
- Execute recalibration according to the calibration interval required in this package insert.

Quality Control

- When new lot used, check or edit the quality control information.
- Scan the control barcode, choose corresponding quality control information and execute testing. For specific information on ordering quality controls, refer to the quality control section of the Analyzer Operating Instructions.

Sample Testing

- After successfully loading the sample, select the sample in interface and edit the assay for the sample to be tested and execute testing. For specific information on ordering patient specimens, refer to the sample ordering section of the Analyzer Operating Instructions.

To ensure proper test performance, strictly adhere to Analyzer Operating Instructions.

Calibration

Traceability: This method has been standardized against the Snibe internal reference standard.

Test of assay specific calibrators allows the detected relative light unit (RLU) values to adjust the master curve.

Recalibration is recommended as follows:

- Whenever a new lot of Reagent or Starter 1+2 is used.

- Every 14 days.
- The analyzer has been serviced.
- Control values lie outside the specified range.

Quality Control

Controls are recommended for the determination of quality control requirements for this assay and should be run in singlicate to monitor the assay performance. Refer to published guidelines for general quality control recommendations, for example Clinical and Laboratory Standards Institute (CLSI) Guideline C24 or other published guidelines¹².

Quality control is recommended once per day of use, or in accordance with local regulations or accreditation requirements and your laboratory’s quality control procedures, quality control could be performed by running the PIGF assay:

- Whenever the kit is calibrated.
- Whenever a new lot of Starter 1+2 or Wash Concentrate is used.

Controls are only applicable with MAGLUMI and Biolumi systems and only used matching with the same top seven LOT numbers of corresponding reagents. For each target value and range refer to the label.

The performance of other controls should be evaluated for compatibility with this assay before they are used. Appropriate value ranges should be established for all quality control materials used.

Control values must lie within the specified range, whenever one of the controls lies outside the specified range, calibration should be repeated and controls retested. If control values lie repeatedly outside the predefined ranges after successful calibration, patient results must not be reported and take the following actions:

- Verify that the materials are not expired.
- Verify that required maintenance was performed.
- Verify that the assay was performed according to the package insert.
- If necessary, contact Snibe or our authorized distributors for assistance.

If the controls in kit are not enough for use, please order PIGF (CLIA) Controls (REF: 160201157MT) from Snibe or our authorized distributors for more.

RESULTS

Calculation

The analyzer automatically calculates the PIGF concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in pg/mL. For further information please refer to the Analyzer Operating Instructions.

Interpretation of Results

The expected range for the PIGF assay was obtained by testing 1551 normal pregnant women individuals in China, gave the following expected value:

Gestational weeks	N	5 th percentile (pg/mL)	50 th percentile (pg/mL)	95 th percentile (pg/mL)
10+0-14+6	154	27.7	51.9	124
15+0-19+6	168	66.9	134	295
20+0-23+6	228	116	254	612
24+0-28+6	330	171	478	1129
29+0-33+6	302	111	488	1284
34+0-36+6	197	74.6	272	1025
37+0-delivery	172	53.2	193	904

Results may differ between laboratories due to variations in population and test method. It is recommended that each laboratory establish its own reference interval.

LIMITATIONS

- Results should be used in conjunction with patient’s medical history, clinical examination and other findings.
- If the PIGF results are inconsistent with clinical evidence, additional testing is needed to confirm the result.
- The assay is only used for an aid in the diagnosis of pregnant women.
- Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits which employ mouse monoclonal antibodies^{13,14}. Additional information may be required for diagnosis.
- Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with *in vitro* immunoassays. Patients routinely exposed to animals or animal serum products can be prone to this interference and anomalous values may be observed¹⁵.
- Bacterial contamination or heat inactivation of the specimens may affect the test results.

SPECIFIC PERFORMANCE CHARACTERISTICS

Representative performance data are provided in this section. Results obtained in individual laboratories may vary.

Precision

Precision was determined using the assay, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): duplicates at two independent runs per day for 5 days at three different sites using three lots of reagent kits (n = 180). The following results were obtained:

Sample	Mean (pg/mL) (n=180)	Within-Run		Between-Run		Reproducibility	
		SD (pg/mL)	%CV	SD (pg/mL)	%CV	SD (pg/mL)	%CV
Serum Pool 1	98.435	3.495	3.55	1.686	1.71	5.777	5.87
Serum Pool 2	1205.469	17.125	1.42	7.429	0.62	25.888	2.15
Serum Pool 3	3940.847	45.120	1.14	11.909	0.30	71.172	1.81
Control 1	199.953	5.071	2.54	2.333	1.17	6.861	3.43
Control 2	1183.775	18.055	1.53	9.402	0.79	40.856	3.45

Linear Range

10.0-10000 pg/mL (defined by the Limit of Quantitation and the maximum of the master curve).

Reportable Interval

6.00-10000 pg/mL (defined by the Limit of Detection and the maximum of the master curve).

Analytical Sensitivity

Limit of Blank (LoB) =2.00 pg/mL.

Limit of Detection (LoD) =6.00 pg/mL.

Limit of Quantitation (LoQ) =10.0 pg/mL.

Analytical Specificity

Interference

Interference was determined using the assay, three samples containing different concentrations of analyte were spiked with potential endogenous interferents in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Interference	No interference up to	Interference	No interference up to
Bilirubin	30 mg/dL	HAMA	30 ng/mL
Hemoglobin	500 mg/dL	Rheumatoid factor	1500 IU/mL
Intralipid	1000 mg/dL	ANA	398 AU/mL

Cross-Reactivity

Cross-reactivity was determined using the assay, three samples containing different concentrations of analyte were spiked with potential cross-reactant in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Cross-reactant	No interference up to	Cross-reactant	No interference up to
VEGF165	10000 pg/mL	VEGF-B167	10000 pg/mL
VEGF121	10000 pg/mL	PDGF-AB	10000 pg/mL

High-Dose Hook

No high-dose hook effect was seen for PIGF concentrations up to 80000 pg/mL.

Method Comparison

A comparison of the PIGF assay with a commercially available immunoassay, gave the following correlations (pg/mL):

Number of samples measured: 162

Passing-Bablok: y=1.0019x-0.5091, r=0.985.

The clinical specimen concentrations were between 10.11 and 9962 pg/mL.

REFERENCES

1. Melnicovici C S, Boşca A B, Şuşman S, et al. Vascular endothelial growth factor (VEGF)-key factor in normal and pathological angiogenesis[J]. Rom J Morphol Embryol, 2018, 59(2): 455-467.
2. Maglione D, Guerriero V, Viglietto G, et al. Isolation of a human placenta cDNA coding for a protein related to the vascular permeability factor[J]. Proceedings of the National Academy of Sciences, 1991, 88(20): 9267-9271.
3. Torry D S, Wang H-S, Wang T-H, et al. Preeclampsia is associated with reduced serum levels of placenta growth factor[J]. American Journal of Obstetrics and Gynecology, 1998, 179(6): 1539-1544.
4. Ghosh S K, Raheja S, Tuli A, et al. Combination of uterine artery Doppler velocimetry and maternal serum placental growth factor estimation in predicting occurrence of pre-eclampsia in early second trimester pregnancy: a prospective cohort study[J]. European Journal of Obstetrics & Gynecology and Reproductive Biology, 2012, 161(2): 144-151.
5. Rana S, Schnettler W T, Powe C, et al. Clinical characterization and outcomes of preeclampsia with normal angiogenic profile[J]. Hypertension in Pregnancy, 2013, 32(2): 189-201.
6. Maynard S E, Venkatesha S, Thadhani R, et al. Soluble Fms-like Tyrosine Kinase 1 and Endothelial Dysfunction in the Pathogenesis of Preeclampsia[J]. Pediatric Research, 2005, 57(5 Part 2): 1R-7R.
7. Ghosh S K, Raheja S, Tuli A, et al. Association between placental growth factor levels in early onset preeclampsia with the occurrence of postpartum hemorrhage: A prospective cohort study[J]. Pregnancy Hypertension: An International Journal of Women's Cardiovascular Health, 2012, 2(2): 115-122.
8. Chappell L C, Duckworth S, Seed P T, et al. Diagnostic Accuracy of Placental Growth Factor in Women With Suspected Preeclampsia: A Prospective Multicenter Study[J]. Circulation, 2013, 128(19): 2121-2131.
9. Schmidt M, Dogan C, Birdir C, et al. Altered angiogenesis in preeclampsia: evaluation of a new test system for measuring placental growth factor[J]. Clinical Chemical Laboratory Medicine, 2007, 45(11).
10. Tarnow L, Astrup A S, Parving H. Elevated Placental Growth Factor (PlGF) Predicts Cardiovascular Morbidity and Mortality in Type 1 Diabetic Patients with Diabetic Nephropathy[J]. Scandinavian Journal of Clinical and Laboratory Investigation, 2005, 65(sup240): 73-79.
11. Pilarczyk K, Sattler K J E, Galili O, et al. Placenta growth factor expression in human atherosclerotic carotid plaques is related to plaque destabilization[J]. Atherosclerosis, 2008, 196(1): 333-340.
12. CLSI. Statistical Quality Control for Quantitative Measurement Procedures: Principles and Definitions. 4th ed. CLSI guideline C24. Wayne, PA: Clinical and Laboratory Standards Institute; 2016.
13. Robert W. Schroff, Kenneth A. Foon, Shannon M. Beatty, et al. Human Anti-Murine Immunoglobulin Responses in Patients Receiving Monoclonal Antibody Therapy[J]. Cancer Research, 1985, 45(2):879-885.
14. Primus F J, Kelley E A, Hansen H J, et al. "Sandwich"-type immunoassay of carcinoembryonic antigen in patients receiving murine monoclonal antibodies for diagnosis and therapy[J]. Clinical Chemistry, 1988, 34(2):261-264.
15. Boscato L M, Stuart M C. Heterophilic antibodies: a problem for all immunoassays[J]. Clinical Chemistry 1988;34(1):27-33.

SYMBOLS EXPLANATIONS



Consult instructions for use



Manufacturer



Temperature limit
(Store at 2-8°C)



Use-by date



Contains sufficient for<n> tests



Keep away from sunlight



This way up



In vitro diagnostic medical device



Catalogue number



Batch code



Authorized representative in the European Community



Kit component



CE marking with notified body ID number

MAGLUMI® and Biolumi® are trademarks of Snibe. All other product names and trademarks are the property of their respective owners.

Summary of safety and performance is available at Eudamed.



Shenzhen New Industries Biomedical Engineering Co., Ltd.
No.23, Jinxiu East Road, Pingshan District, 518122 Shenzhen, P.R. China
Tel: +86-755-21536601 Fax: +86-755-28292740



Shanghai International Holding Corp. GmbH (Europe)
Eiffestrasse 80, 20537 Hamburg, Germany
Tel: +49-40-2513175 Fax: +49-40-255726