



MAGLUMI® hs-CRP (CLIA)

INTENDED USE

The kit is an *in vitro* chemiluminescence immunoassay for the high sensitivity quantitative determination of C-reactive protein (CRP) in human serum and plasma using the MAGLUMI series Fully-auto chemiluminescence immunoassay analyzer and Biolumi series Integrated System, and the assay is used for an aid in assessment of individuals with suspected or confirmed risk of cardiovascular disease.

SUMMARY

C-reactive protein (CRP) is an acute phase inflammatory protein. It is characterized by a discoid configuration of five identical non-covalently bound subunits, each 206 amino acids long with a molecular mass of about 23 kDa. CRP is usually synthesized primarily in liver hepatocytes but has also been reported to be synthesized in other cell types such as smooth muscle cells, macrophages, endothelial cells, lymphocytes, and adipocytes¹. CRP plasma levels are known to increase rapidly within 24-72 hours of acute events such as those caused by bacterial infection, burn and trauma^{2,3}. But in acute viral infections, CRP changes within a narrower range than in bacterial infections⁴. Serum CRP is elevated in bacterial meningitis, and resolution of symptoms following treatment with antibiotics is slow in those with the highest CRP levels¹. Acute urinary tract infection is the most common bacterial infection in childhood⁵. The level of systemic inflammation as measured by circulating levels of CRP has been linked to prognosis in patients with atherosclerotic disease⁶. Apart from diagnosis, CRP has been reported to enable prognosis in rheumatoid arthritis RA, juvenile RA, ankylosing spondylitis, reiter's syndrome, soriatric arthropathy, vasculitic syndrome, chrohn's disease, rheumatic fever, familial Mediterranean fever, myocardial infarction, stable and unstable angina and visceral leishmaniasis, and many patients with active systemic lupus erythematosus (SLE) display only modestly elevated or even normal CRP levels during active disease periods. Marked CRP elevation in a patient with SLE indicates accompanying some intercurrent infections³. The potential use of CRP also as a harbinger of Febrile neutropenia (FN) or blood stream infections (BSI) during acute leukemia induction may help clinicians to identify such events in a timely manner and initiate appropriate therapy preemptively⁷. Currently, CRP is used in medical practice to rule out sepsis and bacterial infection in immunocompetent patients⁴. The level of systemic inflammation as measured by circulating levels of CRP has been linked to prognosis in patients with congestive heart failure, atrial fibrillation, myocarditis, aortic valve disease and heart transplantation. Beyond its basic functions as an important inflammatory biomarker and a proatherosclerotic/prothrombotic agent, CRP has also been shown to predict incidence of myocardial infarction (MI), stroke, and sudden death. CRP emerged as a potent risk marker of recurrent events among patients with no or stable coronary artery disease (CAD), and those undergoing coronary revascularization⁸. Changes in serum CRP levels can be a potential predictor of the outcomes for patients with PE (pulmonary embolism). Additionally, this value of change can notify the presence of some subgroups of PE (massive and minor PE)⁹.

TEST PRINCIPLE

Sandwich chemiluminescence immunoassay.

The prediluted sample, buffer, magnetic microbeads coated with CRP monoclonal antibody are mixed thoroughly and incubated, performing a wash cycle after a precipitation in a magnetic field. ABEI labeled with another CRP monoclonal antibody are then added and incubated, reacting to form sandwich complexes. After precipitation in a magnetic field, the supernatant is decanted and then a wash cycle is performed. Subsequently, the Starter 1+2 are added to initiate a chemiluminescent reaction. The light signal is measured by a photomultiplier as relative light units (RLUs), which is proportional to the concentration of CRP present in the sample.

REAGENTS

Kit Contents

Component	Description	100 tests/kit	50 tests/kit	30 tests/kit
Magnetic Microbeads	Magnetic microbeads coated with CRP monoclonal antibody (~2.00 µg/mL) in PBS buffer, NaN ₃ (<0.1%).	2.5 mL	1.5 mL	1.0 mL
Calibrator Low	A low concentration of CRP antigen in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Calibrator High	A high concentration of CRP antigen in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Buffer	Tris-HCl buffer, NaN ₃ (<0.1%).	12.5 mL	7.0 mL	4.8 mL
ABEI Label	ABEI labeled with CRP monoclonal antibody (~41.7 ng/mL) in Tris-HCl buffer, NaN ₃ (<0.1%).	22.5 mL	12.0 mL	7.8 mL
Diluent	PBS buffer, NaN ₃ (<0.1%).	21.5 mL	11.5 mL	7.5 mL
Control 1	A low concentration of CRP antigen (1.00 mg/L) in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Control 2	A middle concentration of CRP antigen (3.00 mg/L) in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Control 3	A high concentration of CRP antigen (20.0 mg/L) in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL

All reagents are provided ready-to-use.

Warnings and Precautions

- For *in vitro* diagnostic use.
- For professional use only.
- Exercise the normal precautions required for handling all laboratory reagents.
- Personal protective measures should be taken to prevent any part of the human body from contacting samples, reagents, and controls, and should comply with local operating requirements for the assay.
- A skillful technique and strict adherence to the package insert are necessary to obtain reliable results.
- Do not use kit beyond the expiration date indicated on the label.
- Do not interchange reagent components from different reagents or lots.
- Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).
- All waste associated with biological samples, biological reagents and disposable materials used for the assay should be considered potentially infectious and should be disposed of in accordance with local guidelines.
- This product contains sodium azide. Sodium azide may react with lead or copper plumbing to form highly explosive metal azides. Immediately after disposal, flush with a large volume of water to prevent azide build-up. For additional information, see Safety Data Sheets available for professional user on request.

Note: If any serious incident has occurred in relation to the device, please report to Shenzhen New Industries Biomedical Engineering Co., Ltd. (Snibe) or our authorized representative and the competent authority of the Member State in which you are established.

Reagent Handling

- To avoid contamination, wear clean gloves when operating with a reagent kit and sample. When handling reagent kit, replace the gloves that have been in contact with samples, since introduction of samples will result in unreliable results.
- Do not use kit in malfunction conditions; e.g., the kit leaking at the sealing film or elsewhere, obviously turbid or precipitation is found in reagents (except for Magnetic Microbeads) or control value is out of the specified range repeatedly. When kit in malfunction conditions, please contact Snibe or our authorized distributor.
- To avoid evaporation of the liquid in the opened reagent kits in refrigerator, it is recommended that the opened reagent kits to be sealed with reagent seals contained within the packaging. The reagent seals are single use, and if more seals are needed, please contact Snibe or our authorized distributor.
- Over time, residual liquids may dry on the septum surface. These are typically dried salts and have no effect on assay efficacy.
- Use always the same analyzer for an opened reagent integral.
- For magnetic microbeads mixing instructions, refer to the Preparation of the Reagent section of this package insert.
- For further information about the reagent handling during system operation, please refer to Analyzer Operating Instructions.

Storage and Stability

- Do not freeze the integral reagents.



130206023M: 100 tests/kit
130606023M: 50 tests/kit
130706023M: 30 tests/kit

- Store the reagent kit upright to ensure complete availability of the magnetic microbeads.
- Protect from direct sunlight.

Stability of the Reagents	
Unopened at 2-8°C	until the stated expiration date
Opened at 2-8°C	6 weeks
On-board	4 weeks

Stability of Controls	
Unopened at 2-8°C	until the stated expiration date
Opened at 10-30°C	6 hours
Opened at 2-8°C	6 weeks
Frozen at -20°C	3 months
Frozen and thawed cycles	no more than 3 times

SPECIMEN COLLECTION AND PREPARATION

Specimen Types

Only the specimens listed below were tested and found acceptable.

Specimen Types	Collection Tubes
Serum	Tubes without additive/accessory, or tubes containing clot activator or clot activator with gel.
Plasma	K2-EDTA, Na-heparin or Li-heparin

- The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. Follow tube manufacturers' instructions carefully when using collection tubes.

Specimen Conditions

- Do not use heat-inactivated samples or grossly hemolyzed/hyperlipidaemia specimens and specimens with obvious microbial contamination.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some serum specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the serum specimen is centrifuged before a complete clotting, the presence of fibrin may cause erroneous results.
- Samples must be free of fibrin and other particulate matter.
- To prevent cross contamination, use of disposable pipettes or pipette tips are recommended.

Preparation for Analysis

- Inspect all specimens for foam. Remove foam with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.
- Frozen specimens must be completely thawed before mixing. Mix thawed specimens thoroughly by low speed vortexing or by gently inverting. Visually inspect the specimens. If layering or stratification is observed, mix until specimens are visibly homogeneous. If specimens are not mixed thoroughly, inconsistent results may be obtained.
- Specimens should be free of fibrin, red blood cells, or other particulate matter. Such specimens may give reliable results and must be centrifuged prior to testing. Transfer clarified specimen to a sample cup or secondary tube for testing. For centrifuged specimens with a lipid layer, transfer only the clarified specimen and not the lipemic material.
- The sample volume required for a single determination of this assay is 10 µL.

Specimen Storage

Specimens removed from the separator, red blood cells or clot may be stored up to 8 hours at 10-30°C or 3 days at 2-8°C, or 6 months frozen at -20°C. Frozen specimens subjected to up to 2 freeze/thaw cycles have been evaluated.

Specimen Shipping

- Package and label specimens in compliance with applicable local regulations covering the transport of clinical specimens and infectious substances.
- Do not exceed the storage limitations listed above.

Specimen Dilution

- Samples, with CRP concentrations above the analytical measuring interval, can be diluted with Diluent either by following automated dilution protocol or manual dilution procedure. The recommended dilution ratio is 1:5. The concentration of the diluted sample must be >20 mg/L.
- For manual dilution, multiply the result by the dilution factor. For dilution by the analyzers, the analyzer software automatically takes the dilution into account when calculating the sample concentration.

PROCEDURE

Materials Provided

hs-CRP (CLIA) assay, control barcode labels.

Materials Required (But Not Provided)

- General laboratory equipment.
- Fully-auto chemiluminescence immunoassay analyzer Maglumi 600, Maglumi 800, Maglumi 1000, Maglumi 2000, Maglumi 2000 Plus, Maglumi 4000, Maglumi 4000 Plus, MAGLUMI X8, MAGLUMI X3, MAGLUMI X6 or Integrated System Biolumi 8000, Biolumi CX8.
- Additional accessories of test required for the above analyzers include Reaction Module, Starter 1+2, Wash Concentrate, Light Check, Tip, and Reaction Cup. Specific accessories and accessories' specification for each model refer to corresponding Analyzer Operating Instructions.
- Please use accessories specified by Snibe to ensure the reliability of the test results.

Assay Procedure

Preparation of the Reagent

- Take the reagent kit out of the box and visually inspect the integral vials for leaking at the sealing film or elsewhere. If there is no leakage, please tear off the sealing film carefully.
- Open the reagent area door; hold the reagent handle to get the RFID label close to the RFID reader (for about 2s); the buzzer will beep; one beep sound indicates successful sensing.
- Keeping the reagent straight insert to the bottom along the blank reagent track.
- Observe whether the reagent information is displayed successfully in the software interface, otherwise repeat the above two steps.
- Resuspension of the magnetic microbeads takes place automatically when the kit is loaded successfully, ensuring the magnetic microbeads are totally resuspended homogenous prior to use.

Assay Calibration

- Select the assay to be calibrated and execute calibration operation in reagent area interface. For specific information on ordering calibrations, refer to the calibration section of Analyzer Operating Instructions.
- Execute recalibration according to the calibration interval required in this package insert.

Quality Control

- When new lot used, check or edit the quality control information.
- Scan the control barcode, choose corresponding quality control information and execute testing. For specific information on ordering quality controls, refer to the quality control section of the Analyzer Operating Instructions.

Sample Testing

After successfully loading the sample, select the sample in interface and edit the assay for the sample to be tested and execute testing. For specific information on ordering patient specimens, refer to the sample ordering section of the Analyzer Operating Instructions.

To ensure proper test performance, strictly adhere to Analyzer Operating Instructions.

Calibration

Traceability: This method has been standardized against the reference material ERM DA474/IFCC.
Test of assay specific calibrators allows the detected relative light unit (RLU) values to adjust the master curve.
Recalibration is recommended as follows:
• Whenever a new lot of Reagent or Starter 1+2 is used.
• Every 28 days.
• The analyzer has been serviced.
• Control values lie outside the specified range.

Quality Control

Controls are recommended for the determination of quality control requirements for this assay and should be run in singlicate to monitor the assay performance. Refer to published guidelines for general quality control recommendations, for example Clinical and Laboratory Standards Institute (CLSI) Guideline C24 or other published guidelines¹⁰.
Quality control is recommended once per day of use, or in accordance with local regulations or accreditation requirements and your laboratory's quality control procedures, quality control could be performed by running the hs-CRP assay:
• Whenever the kit is calibrated.
• Whenever a new lot of Starter 1+2 or Wash Concentrate is used.
Controls are only applicable with MAGLUMI and Biolumi systems and only used matching with the same top seven LOT numbers of corresponding reagents. For each target value and range refer to the label.
The performance of other controls should be evaluated for compatibility with this assay before they are used. Appropriate value ranges should be established for all quality control materials used.

Control values must lie within the specified range, whenever one of the controls lies outside the specified range, calibration should be repeated and controls retested. If control values lie repeatedly outside the predefined ranges after successful calibration, patient results must not be reported and take the following actions:
• Verify that the materials are not expired.
• Verify that required maintenance was performed.
• Verify that the assay was performed according to the package insert.
• If necessary, contact Snibe or our authorized distributors for assistance.

If the controls in kit are not enough for use, please order hs-CRP (CLIA) Controls (REF: 160201494MT) from Snibe or our authorized distributors for more.

RESULTS

Calculation

The analyzer automatically calculates the CRP concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in mg/L. For further information please refer to the Analyzer Operating Instructions.

Interpretation of Results

The expected range for the hs-CRP assay was obtained by testing 588 apparently healthy individuals in China, gave the following expected value:
≤ 5 mg/L (95th percentile).

Results may differ between laboratories due to variations in population and test method. It is recommended that each laboratory establish its own reference interval. The AHA/CDC Scientific Statement concerning inflammation and cardiovascular markers reports that CRP values < 1 mg/L are low risk for cardiovascular disease prediction; values between 1–3 mg/L are average risk for cardiovascular disease prediction; and values > 3 mg/L are high risk for cardiovascular disease prediction¹¹.
• Screening the entire adult population is not recommended.
• Acute coronary syndrome management should not depend on CRP measurements.
• Patients with persistently unexplained CRP levels above 10 mg/L should be evaluated for noncardiovascular etiologies.
• Serial measurements of CRP should not be used to monitor treatment.
• The average of two CRP results, repeated optimally two weeks apart, should be used on metabolically stable patients.

LIMITATIONS

• Increases in CRP levels are non-specific and should not be interpreted without a complete clinical history. It is recommended that a baseline be determined on individual patients for comparison. Estimations of inflammation should be based on the changes in CRP values from multiple measurements and used in conjunction with the values of other cardiac risk indicators (i.e., HDL, cholesterol, etc.).
• Results should be used in conjunction with patient's medical history, clinical examination and other findings.
• If the CRP results are inconsistent with clinical evidence, additional testing is needed to confirm the result.
• Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits which employ mouse monoclonal antibodies^{12,13}. Additional information may be required for diagnosis.
• Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with *in vitro* immunoassays. Patients routinely exposed to animals or animal serum products can be prone to this interference and anomalous values may be observed¹⁴.
• Bacterial contamination or heat inactivation of the specimens may affect the test results.

SPECIFIC PERFORMANCE CHARACTERISTICS

Representative performance data are provided in this section. Results obtained in individual laboratories may vary.

Precision

Precision was determined using the assay, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): duplicates at two independent runs per day for 5 days at three different sites using three lots of reagent kits (n = 180). The following results were obtained:

Sample	Mean (mg/L) (n=180)	Within-Run		Between-Run		Reproducibility	
		SD (mg/L)	%CV	SD (mg/L)	%CV	SD (mg/L)	%CV
Serum Pool 1	2.932	0.125	4.26	0.072	2.46	0.178	6.07
Serum Pool 2	10.061	0.261	2.59	0.112	1.11	0.456	4.53
Serum Pool 3	19.994	0.293	1.47	0.167	0.84	0.425	2.13
Plasma Pool 1	2.997	0.124	4.14	0.072	2.40	0.178	5.94
Plasma Pool 2	9.838	0.265	2.69	0.108	1.10	0.396	4.03
Plasma Pool 3	19.803	0.279	1.41	0.118	0.60	0.438	2.21
Control 1	1.003	0.037	3.69	0.029	2.89	0.066	6.58
Control 2	3.018	0.110	3.64	0.067	2.22	0.165	5.47
Control 3	19.781	0.788	3.98	0.614	3.10	1.320	6.67

Linear Range

0.300-100 mg/L (defined by the Limit of Quantitation and the maximum of the master curve).

Reportable Interval

0.100-500 mg/L (defined by the Limit of Detection and the maximum of the master curve×Recommended Dilution Ratio).

Analytical Sensitivity

Limit of Blank (LoB) =0.010 mg/L.
Limit of Detection (LoD) =0.100 mg/L.
Limit of Quantitation (LoQ) =0.300 mg/L.

Analytical Specificity

Interference

Interference was determined using the assay, three samples containing different concentrations of analyte were spiked with potential endogenous and exogenous interferents in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Interference	No interference up to	Interference	No interference up to
Hemoglobin	2000 mg/dL	Heparin lithium salt	80 IU/mL
Intralipid	3000 mg/dL	Biotin	0.5 mg/dL
Bilirubin	66 mg/dL	Gentamycin sulfate	120 µg/mL
HAMA	40 ng/mL	Cefotaxim	900 µg/mL
ANA	398 AU/mL	Azithromycin	1.2 mg/dL
Rheumatoid Factor	1500 IU/mL	Angiotensin Converting Enzyme Inhibitor	45 µg/mL
Total protein	12 g/dL	Atenolol	10 µg/mL
K2-EDTA	22.75 µmol/mL	Acetylsalicylic Acid	100 mg/dL
Heparin sodium salt	80 IU/mL	Alteplase	30 µg/mL

Cross-Reactivity

Cross-reactivity was determined using the assay, three samples containing different concentrations of analyte were spiked with potential cross-reactant in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Cross-reactant	No interference up to	Cross-reactant	No interference up to
Recombinant Serum Amyloid A (SAA)	1000 mg/dL	Myoglobin	10000 ng/mL
Human Serum Albumin	1000 mg/dL	cTnlI	100 ng/mL
IgG	1000 mg/dL	CK-MB	2000 ng/mL
Transferrin	3000 mg/dL	PCT	500 ng/mL

High-Dose Hook

No high-dose hook effect was seen for CRP concentrations up to 8000 mg/L.

Method Comparison

A comparison of the hs-CRP assay with a commercially available immunoassay, gave the following correlations (mg/L):

Number of samples measured: 206

Passing-Bablok: y=0.9962x+0.0252, t=0.966.

The clinical specimen concentrations were between 0.5 and 95.5 mg/L.

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SYMBOLS EXPLANATIONS

	Consult instructions for use		Manufacturer
	Temperature limit (Store at 2-8°C)		Use-by date
	Contains sufficient for <=> tests		Keep away from sunlight
	This way up		Authorized representative in the European Community
	<i>In vitro</i> diagnostic medical device		Kit component
	Catalogue number		Batch code
	CE marking with notified body ID number		

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Summary of safety and performance is available at Eudamed.



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