



MAGLUMI® T-Uptake (CLIA)

INTENDED USE

The kit is an *in vitro* chemiluminescence immunoassay for the quantitative determination of T-Uptake in human serum and plasma using the MAGLUMI series Fully-auto chemiluminescence immunoassay analyzer and Biolumi series Integrated System, and the assay is used for an aid in evaluating thyroid function.

SUMMARY

Thyroxine (T4) is the main product of thyroid gland which acts to regulate a wide variety of metabolic functions¹. More than 99% of T4 in the blood is bound to thyroxine-binding globulin (TBG) and to a lesser extent thyroxine-binding prealbumin (TBPA) and albumin. A change in TBG concentration can lead to elevated or lowered total T4 concentrations although does not affect the euthyroid level of free T4². Previous studies show that many factors can cause changes in TBG, resulting in changes in total T4, which can lead to erroneous assessments of thyroid function^{3,4}. The T-Uptake assay measures the available thyroxine binding sites of plasma proteins, which can assess the status of thyroxine binding protein. The combined detection of T-Uptake and total T4 can calculate the free thyroxine index (FT4I) from the quotient of total T4 and TBI (T-Uptake assay results). The FT4I can respond to variations in thyroid binding protein and T4 levels, and provide a useful and accurate basis for clinical diagnosis⁵⁻⁷. Thyroid function tests are designed to distinguish hyperthyroidism and hypothyroidism from the euthyroid state. The comparison of the total T4 with the FT4I shows that the latter test discriminates better between normals and abnormals, and can reliably identify the status of thyroid function in all patients⁸.

TEST PRINCIPLE

Modified competitive chemiluminescence immunoassay.

The sample, free T4 antigen and T4 antigen conjugate coated magnetic microbeads are mixed thoroughly and incubated, the free T4 antigen bind to the free binding sites of thyroxine-binding proteins present in the sample. ABEI labeled with anti-T4 monoclonal antibody are then added and incubated, the remaining free T4 antigen competes with T4 antigen immobilized on the magnetic microbeads for a limited number of binding sites on the ABEI-labeled anti-T4 antibody, forming immuno-complexes. After precipitation in a magnetic field, the supernatant is decanted and then a wash cycle is performed. Subsequently, the Starter 1+2 are added to initiate a chemiluminescent reaction. The light signal is measured by a photomultiplier as relative light units (RLUs), which is proportional to the concentration of T-Uptake present in the sample.

REAGENTS

Kit Contents

Component	Description	100 tests/kit	50 tests/kit	30 tests/kit
Magnetic Microbeads	Magnetic microbeads coated with T4 antigen conjugate (~2.00 µg/mL) in PBS buffer, NaH ₂ (<0.1%).	2.5 mL	1.5 mL	1.0 mL
Calibrator Low	A low concentration of TBG antigen in PBS buffer, NaH ₂ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Calibrator High	A high concentration of TBG antigen in PBS buffer, NaH ₂ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Antigen	T4 antigen in PBS buffer, NaH ₂ (<0.1%).	12.5 mL	7.0 mL	4.8 mL
ABEI Label	ABEI labeled with anti-T4 monoclonal antibody (~0.250 µg/mL) in Tris-HCl buffer, NaH ₂ (<0.1%).	12.5 mL	7.0 mL	4.8 mL
Control 1	A low concentration of TBG antigen (0.700 TBI) in PBS buffer, NaH ₂ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Control 2	A high concentration of TBG antigen (1.10 TBI) in PBS buffer, NaH ₂ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
All reagents are provided ready-to-use.				

Warnings and Precautions

- For *in vitro* diagnostic use.
- For professional use only.
- Exercise the normal precautions required for handling all laboratory reagents.
- Personal protective measures should be taken to prevent any part of the human body from contacting samples, reagents, and controls, and should comply with local operating requirements for the assay.
- A skillful technique and strict adherence to the package insert are necessary to obtain reliable results.
- Do not use kit beyond the expiration date indicated on the label.
- Do not interchange reagent components from different reagents or lots.
- Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).
- All waste associated with biological samples, biological reagents and disposable materials used for the assay should be considered potentially infectious and should be disposed of in accordance with local guidelines.
- This product contains sodium azide. Sodium azide may react with lead or copper plumbing to form highly explosive metal azides. Immediately after disposal, flush with a large volume of water to prevent azide build-up. For additional information, see Safety Data Sheets available for professional user on request.

Note: If any serious incident has occurred in relation to the device, please report to Shenzhen New Industries Biomedical Engineering Co., Ltd. (Snibe) or our authorized representative and the competent authority of the Member State in which you are established.

Reagent Handling

- To avoid contamination, wear clean gloves when operating with a reagent kit and sample. When handling reagent kit, replace the gloves that have been in contact with samples, since introduction of samples will result in unreliable results.
- Do not use kit in malfunction conditions; e.g., the kit leaking at the sealing film or elsewhere, obviously turbid or precipitation is found in reagents (except for Magnetic Microbeads) or control value is out of the specified range repeatedly. When kit in malfunction conditions, please contact Snibe or our authorized distributor.
- To avoid evaporation of the liquid in the opened reagent kits in refrigerator, it is recommended that the opened reagent kits to be sealed with reagent seals contained within the packaging. The reagent seals are single use, and if more seals are needed, please contact Snibe or our authorized distributor.
- Over time, residual liquids may dry on the septum surface. These are typically dried salts and have no effect on assay efficacy.
- Use always the same analyzer for an opened reagent integral.
- For magnetic microbeads mixing instructions, refer to the Preparation of the Reagent section of this package insert.
- For further information about the reagent handing during system operation, please refer to Analyzer Operating Instructions.

Storage and Stability

- Do not freeze the integral reagents.
- Store the reagent kit upright to ensure complete availability of the magnetic microbeads.
- Protect from direct sunlight.

Stability of the Reagents	
Unopened at 2-8°C	until the stated expiration date
Opened at 2-8°C	6 weeks
On-board	4 weeks

Stability of Controls	
Unopened at 2-8°C	until the stated expiration date



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130203014M: 100 tests/kit

130603014M: 50 tests/kit

130703014M: 30 tests/kit

Opened at 10-30°C	6 hours
Opened at 2-8°C	6 weeks
Frozen at -20°C	3 months
Frozen and thawed cycles	no more than 3 times

SPECIMEN COLLECTION AND PREPARATION

Specimen Types

Only the specimens listed below were tested and found acceptable.

Specimen Types	Collection Tubes
Serum	Tubes without additive/accessory, or tubes containing clot activator or clot activator with gel.
Plasma	K2-EDTA, K3-EDTA.

- The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. Follow tube manufacturers' instructions carefully when using collection tubes.

Specimen Conditions

- Do not use heat-inactivated samples or grossly hemolyzed/hyperlipidaemia specimens and specimens with obvious microbial contamination.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some serum specimens, especially those from patients receiving anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the serum specimen is centrifuged before a complete clotting, the presence of fibrin may cause erroneous results.
- Samples must be free of fibrin and other particulate matter.
- To prevent cross contamination, use of disposable pipettes or pipette tips are recommended.

Preparation for Analysis

- Inspect all specimens for foam. Remove foam with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.
- Frozen specimens must be completely thawed before mixing. Mix thawed specimens thoroughly by low speed vortexing or by gently inverting. Visually inspect the specimens. If layering or stratification is observed, mix until specimens are visibly homogeneous. If specimens are not mixed thoroughly, inconsistent results may be obtained.
- Specimens should be free of fibrin, red blood cells, or other particulate matter. Such specimens may give reliable results and must be centrifuged prior to testing. Transfer clarified specimen to a sample cup or secondary tube for testing. For centrifuged specimens with a lipid layer, transfer only the clarified specimen and not the lipemic material.
- The sample volume required for a single determination of this assay is 20 µL.

Specimen Storage

Specimens removed from the separator, red blood cells or clot may be stored up to 8 hours at 10-30°C or 8 days at 2-8°C, or 3 months frozen at -20°C.

Frozen specimens subjected to up to 1 freeze/thaw cycle have been evaluated. Specimens must be mixed thoroughly after thawing.

Specimen Shipping

- Package and label specimens in compliance with applicable local regulations covering the transport of clinical specimens and infectious substances.
- Do not exceed the storage limitations listed above.

Specimen Dilution

- Samples for T-Uptake determinations cannot be diluted, as T4 in the blood is present in free and protein-bound forms which are in equilibrium. A change in the concentration of the binding proteins alters this equilibrium and consequently also the binding capacity being measured.

PROCEDURE

Materials Provided

T-Uptake (CLIA) assay, control barcode labels.

Materials Required (But Not Provided)

- General laboratory equipment.
- Fully-auto chemiluminescence immunoassay analyzer Maglumi 600, Maglumi 800, Maglumi 1000, Maglumi 2000, Maglumi 2000 Plus, Maglumi 4000, Maglumi 4000 Plus, MAGLUMI X3, MAGLUMI X6, MAGLUMI X8 or Integrated System Biolumi 8000 and Biolumi CX8.
- Additional accessories of test required for the above analyzers include Reaction Module, Starter 1+2, Wash Concentrate, Light Check, Tip, and Reaction Cup. Specific accessories and accessories' specification for each model refer to corresponding Analyzer Operating Instructions.
- Please use accessories specified by Snibe to ensure the reliability of the test results.

Assay Procedure

Preparation of the Reagent

- Take the reagent kit out of the box and visually inspect the integral vials for leaking at the sealing film or elsewhere. If there is no leakage, please tear off the sealing film carefully.
- Open the reagent area door; hold the reagent handle to get the RFID label close to the RFID reader (for about 2s); the buzzer will beep; one beep sound indicates successful sensing.
- Keeping the reagent straight insert to the bottom along the blank reagent track.
- Observe whether the reagent information is displayed successfully in the software interface, otherwise repeat the above two steps.
- Resuspension of the magnetic microbeads takes place automatically when the kit is loaded successfully, ensuring the magnetic microbeads are totally resuspended homogenous prior to use.

Assay Calibration

- Select the assay to be calibrated and execute calibration operation in reagent area interface. For specific information on ordering calibrations, refer to the calibration section of Analyzer Operating Instructions.
- Execute recalibration according to the calibration interval required in this package insert.

Quality Control

- When new lot used, check or edit the quality control information.
- Scan the control barcode, choose corresponding quality control information and execute testing. For specific information on ordering quality controls, refer to the quality control section of the Analyzer Operating Instructions.

Sample Testing

- After successfully loading the sample, select the sample in interface and edit the assay for the sample to be tested and execute testing. For specific information on ordering patient specimens, refer to the sample ordering section of the Analyzer Operating Instructions.

To ensure proper test performance, strictly adhere to Analyzer Operating Instructions.

Calibration

Traceability: This method has been standardized against the Snibe internal reference standard.

Test of assay specific calibrators allows the detected relative light unit (RLU) values to adjust the master curve.

Recalibration is recommended as follows:

- Whenever a new lot of Reagent or Starter 1+2 is used.
- Every 28 days.
- The analyzer has been serviced.
- Control values lie outside the specified range.

Quality Control

Controls are recommended for the determination of quality control requirements for this assay and should be run in singlicate to monitor the assay performance. Refer to published guidelines for general quality control recommendations, for example Clinical and Laboratory Standards Institute (CLSI) Guideline C24 or other published guidelines⁹.

Quality control is recommended once per day of use, or in accordance with local regulations or accreditation requirements and your laboratory's quality control procedures, quality control could be performed by running the T-Uptake assay:

- Whenever the kit is calibrated.
- Whenever a new lot of Starter 1+2 or Wash Concentrate is used.

Controls are only applicable with MAGLUMI and Biolumi systems and only used matching with the same top seven LOT numbers of corresponding reagents. For each target value and range refer to the label.

The performance of other controls should be evaluated for compatibility with this assay before they are used. Appropriate value ranges should be established for all quality control materials used.

Control values must lie within the specified range, whenever one of the controls lies outside the specified range, calibration should be repeated and controls retested. If control values lie repeatedly outside the predefined ranges after successful calibration, patient results must not be reported and take the following actions:

- Verify that the materials are not expired.
- Verify that required maintenance was performed.
- Verify that the assay was performed according to the package insert.
- If necessary, contact Snibe or our authorized distributors for assistance.

If the controls in kit are not enough for use, please order T-Uptake (CLIA) Controls (REF: 160201180MT) from Snibe or our authorized distributors for more.

RESULTS

Calculation

The analyzer automatically calculates the T-Uptake concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in TBI. For further information please refer to the Analyzer Operating Instructions. In addition, the combined detection of T-Uptake and total T4 can calculate the free thyroxine index (FT4I) with the quotient of total T4 and TBI (T-Uptake assay results).

Interpretation of Results

The expected range for the T-Uptake assay was obtained by testing 252 apparently healthy individuals in China, gave the following expect value: 0.8-1.3 TBI (2.5th-97.5th percentiles).

FT4I (free thyroxine index):

The expected range for the FT4I (the quotient of MAGLUMI Total T4 and TBI) was obtained by testing 252 apparently healthy individuals in China, gave the following expect value: 50-120 ng/mL (2.5th-97.5th percentiles).

Results may differ between laboratories due to variations in population and test method. It is recommended that each laboratory establish its own reference interval.

LIMITATIONS

- Results should be used in conjunction with patient's medical history, clinical examination and other findings.
- If the T-Uptake results are inconsistent with clinical evidence, additional testing is needed to confirm the result.
- The assay is mainly used to evaluate individuals with suspected or confirmed abnormal thyroid function.
- The test cannot be used in patients receiving treatment with lipid lowering agents containing D-T4. If the thyroid function is to be checked in such patients, the therapy should first be discontinued for 4-6 weeks to allow the physiological state to become re-established¹⁰.
- Autoantibodies to thyroid hormones can interfere with the assay. Binding protein anomalies seen with FDH (familial dysalbuminemic hyperthyroxinemia), for example, may cause values which, while characteristic of the condition, deviate from the expected results¹¹.
- Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits which employ mouse monoclonal antibodies^{12,13}. Additional information may be required for diagnosis.
- Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with *in vitro* immunoassays. Patients routinely exposed to animals or animal serum products can be prone to this interference and anomalous values may be observed¹⁴.
- Bacterial contamination or heat inactivation of the specimens may affect the test results.

SPECIFIC PERFORMANCE CHARACTERISTICS

Representative performance data are provided in this section. Results obtained in individual laboratories may vary.

Precision

Precision was determined using the assay, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute); duplicates at two independent runs per day for 5 days at three different sites using three lots of reagent kits (n = 180). The following results were obtained:

Sample	Mean (TBI) (n=180)	Within-Run		Between-Run		Reproducibility	
		SD (TBI)	%CV	SD (TBI)	%CV	SD (TBI)	%CV
Serum Pool 1	0.307	0.012	3.91	0.007	2.28	0.018	5.86
Serum Pool 2	0.839	0.030	3.58	0.008	0.95	0.040	4.77
Serum Pool 3	1.703	0.052	3.05	0.040	2.35	0.083	4.87
Plasma Pool 1	0.315	0.010	3.17	0.007	2.22	0.014	4.44
Plasma Pool 2	0.833	0.029	3.48	0.016	1.92	0.041	4.92
Plasma Pool 3	1.747	0.053	3.03	0.025	1.43	0.095	5.44
Control 1	0.694	0.027	3.89	0.021	3.03	0.043	6.20
Control 2	1.104	0.044	3.99	0.020	1.81	0.058	5.25

Linear Range

0.200-2.00 TBI (defined by the Limit of Quantitation and the maximum of the master curve).

Reportable Interval

0.150-2.00 TBI (defined by the Limit of Detection and the maximum of the master curve).

Analytical Sensitivity

Limit of Blank (LoB) =0.100 TBI.

Limit of Detection (LoD) =0.150 TBI.

Limit of Quantitation (LoQ) =0.200 TBI.

Analytical Specificity

Interference

Interference was determined using the assay, three samples containing different concentrations of analyte were spiked with potential endogenous and exogenous interferences in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Interference	No interference up to	Interference	No interference up to
Bilirubin	41 mg/dL	Sodium salicylate	50.0 mg/dL
Hemoglobin	2000 mg/dL	Aspirin	50.0 mg/dL
Intralipid	2000 mg/dL	Ibuprofen	50.0 mg/dL
Biotin	5 mg/dL	Paracetamol	20.0 mg/dL
HAMA	40 ng/mL	Phenytoin	5.0 mg/dL
Rheumatoid factor	1500 IU/mL	Amiodarone	20.0 mg/dL
ANA	398 AU/mL	Propylthiouracil	30.0 mg/dL
Phenylbutazone	15.0 mg/dL	-	-

Cross-Reactivity

Cross-reactivity was determined using the assay, three samples containing different concentrations of analyte were spiked with potential cross-reactant in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Cross-reactant	No interference up to	Cross-reactant	No interference up to
3-Iodo-L-tyrosine	10000 µg/dL	3,3',5'-Triiodo-L-thyronine	200 µg/dL
3,5-Diiodo-L-tyrosine	1000 µg/dL	3,3',5'-Triiodothyroacetic acid	10 µg/dL
3,5-Diiodo-L-thyronine	250 µg/dL	3,3',5',5'-Tetraiodothyroacetic acid	10 µg/dL
3,3',5'-Triiodo-L-thyronine	5 µg/dL	-	-

High-Dose Hook

No high-dose hook effect was seen for sample concentrations up to 12.0 TBI.

Method Comparison

A comparison of the T-Uptake assay with a commercially available immunoassay, gave the following correlations (TBI):

Number of samples measured: 120

Passing-Bablok: y=1.0053x-0.0012, r=0.977.

The clinical specimen concentrations were between 0.211 and 1.89 TBI.

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SYMBOLS EXPLANATIONS

	Consult instructions for use		Manufacturer
	Temperature limit (Store at 2-8°C)		Use-by date
	Contains sufficient for<n> tests		Keep away from sunlight
	This way up		Authorized representative in the European Community
	<i>In vitro</i> diagnostic medical device		Kit component
	Catalogue number		Batch code
	CE marking with notified body ID number		

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