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# العدوى الطفيلية الأولية التي تصيب الأسماك والمشاكل الصحية الناتجة عن تفشي الأمراض الطفيلية

#### الملخص:

الأسماك هي مصدر حيوي لغذاء الإنسان يعتبر من أهم مصادر البروتينات عالية الجودة للإنسان، حوالي 16% من البروتين الحيواني الذي يتم استهلاكه من قبل سكان العالم. العديد من مشاكل الأسماك، سواءً كانت في المزارع السمكية أو في الطبيعة فإنها ناتجة عن الطفيليات الأولية المختلفة. أهم طفيليات الأوليات التي تصيب Trichodina sp, Myxobolus sp Tetrahymena sp, Ichthyobodo necator الأسماك مثل Cryptopia spp, Haemogregarina spp, Trypanosoma spp 'Chilodonella sp 'Chilodonella sp 'Chilodonella sp 'Chilodonella sp 'Chilodonella sp 'Chilodonella sp 'Chilodonella spp Ichthyophthirius multifiliis Henneguya, spp Microsporidia spp الطفيليات الأولية خسائر فادحة في أحواض الأسماك فهي تمثل أحد التهديدات الخطيرة على صحة الأسماك بعد تقشي المرض، التعامل معقدًا مع المرض وإلى حد كبير يعتمد على اكتشاف العوامل الممرضة ومضيفيها وانتشارها وعلاجها والوقاية منها وإدارة الصحة العامة. تهدف هذه المراجعة إلى تسليط الضوء على العدوى الطفيلية الأولية للأسماك بالإضافة إلى معرفة المشكلات الصحية الناتجة عن تقشى الأمراض الطفيلية.

### الكلمات المفتاحية: العدوي الطفيلية الأولية، أمراض الأسماك.

#### **Abstract**:

Fish is one of the most important sources of animal protein, it is a source of high-quality protein, and it provides 16% of the animal protein consumed by the world's population. Many fish disasters, both in nature and fish farms were caused by different protozoa parasites. Some most important protozoan parasites in fish such as *Trichodina sp*, *Tetrahymena sp.*, *Chilodonella sp*, *Myxobolus spp*, *Ichthyophthirius multifiliis*, *Ichthyobodo necator*, *Haemogregarina spp*. *Trypanosoma spp*. *Cryptopia spp*, *Microsporidia spp*, *Henneguya spp*, *Thelohania spp*.Protozoan parasites cause dangerous losses in fishponds and wild in fish and occupy a very important sector as one of the serious threats to fish health after outbreak of disease. Disease manipulate is complex and relies largely on pathogen detection, their hosts, prevalence, treatment, prevention, and public health management. The aim of this review is highlight on protozoan parasitic infection of Fish in addition to knowing, health problems resulting from outbreaks of parasitic diseases.

**Keywords:** Protozoan Parasitic Infection, Fish diseases.

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#### Introduction

Fish is a vital source of food for humans. It is the most important source of high-quality protein for humans, providing 16% of animal protein consumed by the world's population (Bilqees *et al*, 2003). Parasites are an important collection of pathogen reasons infection and sicknesses for fishes in freshwater and marine environments. With the increasing interests in aquaculture parasitic infections are getting threats for fish health management and aquatic crop production in the world. (Areerat *et al*, 1981). Protozoans are widespread parasites of both freshwater and marine fishes. To date, around 2420 species in 6 phyla of the protozoa group have been egistered infecting fish; more than 800 are found in marine fish (Lorn and Dykova, 1992).

The Phylum Protozoa gathers numerous organisms evolutionarily different which could act as ecto and endoparasites in fish (lom & dykova, 1992). Protozoa's are single-celled organisms, numerous of which are free-living in the aquatic environment. Typically, no intermediate host is needed for the parasite to reproduc (direct life cycle) (Ryan *et al*, 2019). In general protozoa are one of the main sectors of fish parasites that have been long overlooked due to its inherent difficulty instudying in comparison to other large parasites (Omeji *et al*, 2011).

Protozoans vary in form and size and live mostly on the gills, fins, and skin of fish. Nevertheless some, *including Cryptobia, Hexamita*, and *Myxozoa*, live in the internal organs (Roberts and Stepherd, 1997). *Apiosoma, Chilodonella, Balantidium, Epistylis, Ichthyophthirius multifiliis, Rhynchodinium paradoxum, Nyctotherus, Tetrahymena* and *Trichodinidae* are the major representatives. (EL-Tantawy *et al*, 2013). *Trichodina spp, Ichthyobodo necator* and *Ichthyophthirius multifiliis* are protozoan parasites that cause fish deaths (Durborow, 2003).

Parasitic illnesses are one of the most severe problems in fishes, though not of much concern among the wild fish stock because in most cases, no significant harm appears to have been done to them (Jithendran, 2014). Therefore, there are simplest. little reports of parasites causing death or intense damage to the fish populations, however this could be in large part due to the fact such effects pass unnoticed (Roberts, 2001). However, in times in which hosts are overcrowded consisting of in aquaria or in fish farms, parasitic illnesses can spread very unexpectedly causing excessive mortality (Imam and Dewu, 2010). Parasites are able to inflicting harm to the fish, either via damage to the tissues or organs in the process of burrowing or eating meals or the removal of digested food inside the gut of the fish in addition to the secretion of proteolytic enzymes (Moratal *et al*, 2020). Control of fish parasites requires knowledge of the parasites, their hosts, and their prevalence and distribution (Mitchum, 1995). The aim of this review is highlight on protozoan parasitic infection of fish in addition to knowing, health problems resulting from the spread of parasitic diseases.

### protozoan parasitic infection

Most of the typically encountered fish parasites are protozoans. They are uncellular organisms, Lots of which are free-living within the aquatic surroundings. Their potential to multiply on or within their hosts makes them often very risky to fish (Chandra, 2004). Ectoparasitic ciliates represent one of the most hazardous threats to fish health (Pádua *et al.*, 2013). The ciliary protozoan, Ichthyophthirius multifiliis (Foquet), the causative agent of Ich or ichthyophthiriasis, is one of the maximum important pathogenic protozoan

parasites of cultured fish (Schäperclaus, 1991). And wild fish (Francis-Floyd & Reed ,1991). In general, the theronts foray, and penetrate the dermis of the fish and form cavities enclosed via epithelial cells where they settle and feed. This method is mediated through a combination of mechanical and enzymatic moves (Wang et al., 2019). This process is mediated by a combination of mechanical and enzymatic actions (Wang et al., 2019). Ichthyophthirius multifiliis was the most common protozoan parasites found in Clarias gariepinus. These protozoan parasites comprise (37.08%) of the total parasites found in fish that live in the pond, and (42.51%) of wild fish (Omeji et al,2011). Trichodina, a genus of ciliate protists, belongs to the family Trichodinidae and is well known as the causative agent of trichodiniasis in numerous aquatic animals, especially both cultured and wild fish (Marcotegui, et al, 2018). The activities of Trichodina in infected fish damage epithelial cells and make pressure in the infected fish. Therefore, medical symptoms ininfected host vary from deat to asphyxia. (Valladao et al., 2014). Those ectoparasites are regarded to be the motive of dying of mas fish larvae (Cyprinus carpio), mujair (Oreochromis mossambicus), tawes (Puntius javanius), nila (Oreochromis niloticus), betutu (Oxyeleotris marmorata), rainbow trout (Salmo gairdneri), salmon (Salmo salar), and gurami fish (Osphronemus gourami) (Anisah et al., 2017). Chilodonella spp. is a cilium parasite that parasitizes integument and gills of several fresh water fish species (Pavanelli et al, 2008). and cause severe epizootic outbreaks in wild and farmed freshwater fishes (Lom & Dykova, 1992). Chilodonella is belonging to that may reason infected fish to secrete excessive mucus. sobkingdom a protozoa, Infected fish may flash and display similar clinical signs of inflammation and lots of fish die when infections turn out to be temperate (Abdel-Baki et al., 2014). chilodonella species recorded from gills of Oreochromus niloticus and Clarias gariepinus (Hossin, 1992). Tetrahymena spp. are ubiquitous free-living hymenostome ciliates, that cause disease in a wide variety of fish, crustaceans, amphibians and turbellarians (Ponpornpisit et al, 2000). Tetrahymenosis has been reported in different species of ornamental and edible fish (Astrofsky et al., 2002). Pathogenic characteristics of infected shes by Tetrahymena spp. Pathogenic characteristics of infected shes via Tetrahymena spp. Are analogous, : owning whitish lesions on the body surface, and affected organs involve, the pores and viscera, skin, musculature, eye socket and spinal cord; masses of ciliates can be detected in copious amount of mucus and between spaces in the damaged tissues (Lawhavinit et al., 2002).

Ichthyobodo necator is An ectoparasite that belongs flagellate and infects the gills and skin of fishes. Flagellates identified as *I. necator* have repeatedly been implicated in diseases and mortalities among ornamental and farmed fishes (Woo,1994). variants have also been observed on captive marine fish (Beck et al, 1996), The parasite has been found in the Pacific Ocean, Atlantic Ocean, and adjacent seas including Australian waters (Ueki et al, 1998) Large numbers of *I. necator* trophozoites can destroy the epithelial layer of the skin and gills, which in turn disrupts homeostasis, causing osmoregulatory stress and death (Noga, 1996).

*Cryptobia* species are flagellated protists, distantly associated with Spironucleus and Hexamita, however not almost as well understood. Like Hexamita and Spironucleus, Cryptobia spp. Are very small, (single-celled) organism and, therefore, may be difficult to identify and observed. There were 52 species of *Cryptobia* identified in fish (Floyd & Yanong, 1999). *Cryptobia* attachment via the flagellum does no induce result in any

pathological or even ultrastructural cellular damage (Lom, 1986). Contrary to reports of morbidities related to this parasite (Woo, 1987) Despite the fact that there are some of reports on bad condition and mortalities, specially of fry (Lightner *et al*, 1988).

The genus Myxobolus (family Myxobolidae) is certainly one of the myxosporean groups and its members are significant pathogens of marine fish and freshwater in numerous geographical areas (Lom & Dyková, 2013). The traditionally classified with the protozoa due to their small size, it is now recognised that myxozoans are degenrate metazoans ,transmission is via an oligochaete alternate host. The Myxosporea are relationship to the Myxozoa, a class of microscopic parasites. Species have been described from various tissues and organs, including gills (Azevedo et al, 2014), liver (Svobodova & Kolarova, 2004). Species have been described from various tissues and organs, including gills (Azevedo et al, 2014), liver (Carriero et al, 2013). Among the myosporean, species of the genus myxobolus are, so, far the fo oremostunre markably found in fish, with about 856 known species throughout the world (Eiras et al ,2005). of them, about 12 species were described from tilapias spp (abdel-azeem et al, 2015) The most common symptoms of the disease are weakness, emaciation, scale protrusion, loss of scales, abnormal pigmentation Spores released from the infected and dead fishes remain viable for quite a long period in the pond bottom before they infect new hosts. (Kumar, 1992).

The genus Henneguya Thélohan, 1892, in family Myxobolidae, is the third most speciose myxosporean genus after Myxidium and Myxobolus, with > 200 Henneguya species described (Székely et al, 2018) The type species, H. psorospermica was described by Thelohan(1895) from the gills of both pike and perch (Perca fluviatilis L.). Of them the pike, mentioned first by Thélohan, should be regarded as type host. Other pike-infecting Henneguya species were recorded from the gills (H. lobosa (Cohn, 1895). (Biance et al, 2003) described a new species of Myxosporea, called Henneguya chydadea, is parasitizing the gills of fish collected from a lake in state of Säo Paulo, Brazil. They recorded that about (88.3%) of examined fish had gills parasitized by Myxosporeans. The spread of the parasite ranged about (80%) inside the fall and spring seasons, (93%) during the summer season and (100%) during the winter season. Infections can purpose mortalities in fishes, if parasites proliferate to high severity on the gills and reason breathing failure,, particularly in juvenile fish (Whitaker et al, 2005).

Microsporidia are common pathogens of numerous aquatic organisms, including crustaceans and amphipods, and members from some 18 genera of these parasites have been described in fishes (Lom & Nilsen, 2003). Among the microsporidian genera infecting species of freshwater crayfish, Thelohania is a serious pathogen in many countries (Evans & Edgerton, 2002). The first 'true' Thelohania with gene sequence data available, *T. butleri*, was identified from Canadian pink shrimp (Pandalus jordani) off the coast of British Columbia, Canada (Brown & Adamson, 2006). A dangerous protozoan pathogen overwhelmingly infests the black tiger shrimp, that leads to the cotton illness. *Thelohania sp.* is secluded from striated muscles, hepatopancreas, and intestine; these organs often degenerate (Prasertpol, 1989).

Haemogregarines are adeleorine that belonging to apicomplexan protists and live in the blood cells and tissues of differents of vertebrates and are especially prevalent in

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marine fishes (Davies *et al*, 2004). Knowledge of marine fish haematozoans from South Africa is limited. Two species of haemogregarines, Haemogregarina (sensu lato) *bigemina* (Laveran & Mesnil, 1901) and Haemogregarina (sensu lato) *koppiensis* (Smit & Davies, 2005). Haemogregarina(sensu lato) bigemina Laveran and Mesnil (1901) having first been defined from intertidal blenniid fishes Lipophrys pholis and Coryphoblennius galerita (Linnaeus, 1758) in northern France in 1901 might be the most enigmatic of the marine fish haemogregarines (Laveran & Mesnil,1901). (Davies *et al*, 2004) reviewed past and current researches on the Haemogregarina bigemina, recorded from (96) species of fishes at Mesnil. The parasite undergoes intraerythrocytic binary fission finally forming mature paired gamonts.

Trypanosoma species are almost all heteroxenous and parasites of the blood of all classes of vertebrates, including marine and freshwater teleost and elasmobranch fish all over the world (Hayes et al, 2014). About 184 species that belonging Trypanosomatids in the genus Trypanosoma assigned to infect fish .Trypanosoma have a single flagellum and a single disc shaped kinetoplast. Infections in most species of fish are transmitted by leeches (Paperna, 1996). Despite most infected fish without clinical signs, fish trypanosomiasis can be acute at heavy parasitemias, and clinical signs include anaemia, leukocytosis, hypoglycemia and splenomegaly (Su et al, 2014). The haemoparasite belonging to the Genus Trypanosoma species were reported to appear, in Lake Victoria by way of Paperna (1996) in Esculenta (50%), Oreochromis variabilis (54%), O. Clarias gariepinus and Bagrus spp (Paperna, 1996).

Protozoa are common tropical and subtropical marine water fish parasites that affect public health and cause losses to fishes (Ahmed *et al*, 2021). In Libya Coasts (Ahmed *et al*, 2021) examined Mediterranean horse macherel (*Trachurus mediterraneus*) from Zliten coast were identified 7 species of parasitic protozoan particularly *Haemogregarina spp.*, *Cryptopia spp Trypanosoma spp.*, *Microsporidia spp.*, *Henneguya spp.*, *Thelohania spp. and Ichthyophthirius multifiliis*. Among the examined fish, the high desnsity of parasitic infectious in internal organ was observed in kideny (0.375), liver (0.5), blood and spleen (0.25), respectively. in addation (SH & & RA, 2022) made general survey of protozoan parasites that infect Sawrow fish (*Trachurus mediterraneus*) from Zliten coast, Libya, The outcomes confirmed positive impact of *haemogregarina* inside the Sawrow Blood registered, highest dispersal and abundance amidst all of the different organs in Sawrow fish observed kideny, liver and spleen respectively.

(younis, 2012) made survey of protozoan parasites that infect *Tilapia zillii* (Gervais, 1852) and *Mugil cephalus* (Linnaeus, 1758) from Ain Ziana lagoon, Benghazi, Libya. the results confirmed that six species of ectoprotozoan parasites had been in the course of the examination of gills and skin these fishes. Theses parasites were *Tetrahymena sp, Chilodonella sp,Trichodina sp,Ichthyophthirius multifiliis Ichthyobodo necator, Myxobolus spp.* 

#### **Conclusions**

Fish is important to human; it is of importance in the diet of different countries. Infectious fish illnesses result from fungi, viruses, micro organism, and parasites. Parasites that affect fish include protozoans and metazoan. parasitic protozoans that affect fish include *Trichodina sp, Tetrahymena sp.*, *Chilodonella sp, Myxobolus spp*, *Ichthyophthirius multifiliis, Ichthyobodo necator, Haemogregarina spp. Trypanosoma spp. Cryptopia spp, Microsporidia spp, Henneguya spp, Thelohania spp. Ichthyophthirius* 

*multifiliis*. Parasitic infestation repeatedly occurs in fish that lead retarded growth rate, consumer rejection, reduced production, low reproduction and collective mortality in fish. Parasites affect on the fish health through the affect the skin and gills of fishes. And through the tissues or organs damage and the appears of disease.

To overcome the consequences of parasitic infection on fish and public health, it's important to behave upon each health constraint based totally on scientifically verified and advocated in addition to locally applicable methods and additionally epidemiological procedures needed in maintaining aquatic animal fitness safe.

#### **Reference:**

**Abdel-Baki**, A. A. S., & Al-Quraishy, S. (2014). First record of Chilodonella spp.(Ciliophora: Chilodonellidae) in cultured Nile tilapia (Oreochromis niloticus) in the central region of Saudi Arabia. Pakistan Journal of Zoology, 46(3)

**Abdel-Baki**, A. A. S., Zayed, E., Sakran, T., & Al-Quraishy, S. (2015). A new record of Myxobolus brachysporus and M. israelensis in the tilapia (Oreochromis niloticus) collected from the Nile River, Egypt. *Saudi Journal of Biological Sciences*, 22(5), 539-542.

**Ahmed**, S. H., Yousif, R. A., Eldehidya, R., & Elaakary, S. (2021). Investigation of Haemogregarina in Sawrow fish (Trachurus mediterraneus) in Zliten coastal area, Libya. **Anisah**, N., Rokhmani, R., & Riwidiharso, E. (2017). Intensitas dan variasi morfometrik Trichodina sp. pada benih ikan gurami (Osphronemus gouramy Lacepede) pendederan i yang dijual di pasar ikan Purwonegoro Kabupaten Banjarnegara. Majalah Ilmiah Biologi BIOSFERA: A Scientific Journal, 33(3), 134-141

**Astrofsky**, K. M., Schech, J. M., Sheppard, B. J., Obenschain, C. A., Chin, A. M., Kacergis, M. C., ... & Fox, J. G. (2002). High mortality due to Tetrahymena sp. infection in laboratory-maintained zebrafish (Brachydanio rerio). *Comparative medicine*, *52*(4), 363-367.

**Areerat**, S., Boonyaratpalin, B. S., Chinabut, S., Pawaputanon, K., Macrae, I. H., Muir, J. F., ... & Sommerville, C. (1981). A Handbook of Diseases of Cultured Clarias (Pla Duk) in Thailand. National Inland Fisheries Institute. *Freshwater Fisheries Division*, *Department of Fisheries, Thailand*, 59.

**Azevedo**, R. K., Vieira, D. H. M. D., Vieira, G. H., Silva, R. J., Matos, E., & Abdallah, V. D. (2014). Phylogeny, ultrastructure and histopathology of Myxobolus lomi sp. nov., a parasite of Prochilodus lineatus (Valenciennes, 1836)(Characiformes: Prochilodontidae) from the Peixes River, São Paulo State, Brazil. *Parasitology international*, 63(2), 303-307.

**Beck**, K., Lewbart, G., & Piner, G. (1996). The occurrence of an Ichthyobodo-like organism on captive Atlantic spadefish, Chaetodipterus faber (Broussonet). *Journal of Fish Diseases*, 19(1), 111-112.

**Bianca** B.; Nelson S. and Sarah A. (2003): "A new species of Henneguya, a Gill parasite of Astyanax altiparanae (pisces: Characidae) from Brazil, with comments on Histopathology and seasonality", Mem. Inst. Oswaido Cruz. Riode Janeiro, Vol. 98, No. (6), PP. 761 – 765.

**Bilqees**, F.M., Khatoon, N., & Hadi, R. (2003). A new species of the genus Stromaturus, from a fish, Otolithus argenteus of Karachi coast. India. J.Exp.Zool. 8, 2:435-9.

Brown, A. M., & Adamson, M. L. (2006). Phylogenetic distance of Thelohania butleri

Johnston, Vernick, and Sprague, 1978 (Microsporidia; Thelohaniidae), a parasite of the smooth pink shrimp Pandalus jordani, from its congeners suggests need for major revision of the genus Thelohania Henneguy, 1892. *Journal of Eukaryotic Microbiology*, 53(6), 445-455

**Carriero**, M. M., Adriano, E. A., Silva, M. R., Ceccarelli, P. S., & Maia, A. A. (2013). Molecular phylogeny of the Myxobolus and Henneguya genera with several new South American species. *PLoS One*, 8(9), e73713.

**Chandra**, K. J. (2004). Fish parasitology. K. R. Choudhury, 34/A/2, *Ram Babu Road*, *Mymensingh*-196.

Cohn, L. (1895). Über die Myxosporidien von esox lucius und perta fluviatilis.

**Davies**, A. J., Smit, N. J., Hayes, P. M., Seddon, A. M., & Wertheim, D. F. (2004). Haemogregarina bigemina (Protozoa: Apicomplexa: Adeleorina)—past, present and future. *Folia Parasitologica*, 51(2/3), 99-108.

**Durborow**, R. (2003). Protozoan Parasites . SRAC Publication No 4701. Mississippi State University, Mississippi, *Starkville*, USA.

**Eiras**, J. C., Molnár, K., & Lu, Y. (2005). Synopsis of the species of Myxobolus bütschli, 1882 (Myxozoa: Myxosporea: Myxobolidae). Systematic parasitology, 61(1), 1-46.

**El-Tantawy**, S. A. M., Reda, E. S. A., Abdel-Aziz, A., Abou El-Nour, M. F., & Rady, I. (2013). Apiosoma spp. and Scopulata epibranchialis infesting Nile perch fish Lates niloticus in Dakahlia Province, Egypt. *NY Sci. J*, 6(6), 111-118.

**Evans,** L. & Edgerton, B. F. (2002). Pathogens parasites and commensals. In: Holdich D.M.(ed.), Biology of Freshwater Crayfish, *Blackwell Science, Oxford*, 377–438. **Floyd**, R.F., & Yanong, R. (1999). Cryptobia iubilans in Cichlids. The Institute of Food and Agricultural Sciences (*IFAS*) *IFAS Extension*.

**Francis-Floyd**, R., & Reed, P. (1991). Ichthyophthirius multifiliis (white spot) infections in fish. Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida.

**Hayes**, P. M., Lawton, S. P., Smit, N. J., Gibson, W. C., & Davies, A. J. (2014). Morphological and molecular characterization of a marine fish trypanosome from South Africa, including its development in a leech vector. *Parasites & vectors*, 7(1), 1-11.

**Hossin**, M. M. (1992). Studies on some gill affections in freshwater fishes. MV Sc. Thesis, Fac. Of Vet. Med., *Beni Sueif*, Cairo University.

**Imam**, T. S., & Dewu, R. A. (2010). Survey of piscine ecto-and intestinal parasites of clarias species sold at Galadima Road Fish Market, Kano metropolis, Nigeria. *Bioscience Research Communications*, 22(4), 209-214.

**Jithendran**, K. P. (2014). Parasites and Parasitic Diseases in Fish Culture System. Not Available

**Kumar**, D. (1992). Fish culture in undrainable ponds: A manual for extension.

**Laveran**, A., & Mesnil, F. (1901). Deux hémogrégarines nouvelles des poissons. *CR Acad. Sci. Paris*, 133, 572-577.

**Lawhavinit**, O. A., Chukanhom, K., & Hatai, K. (2002). Effect of Tetrahymena on the occurrence of achlyosis in the guppy. *Mycoscience*, 43(1), 27-31.

**Ligthner**, D., Redman, R., Mohney, L., Dickenson, G., & Fitzsimmons, K. (1988). Major disease encountered in controlled environment culture of tilapias in fresh-and brackishwater over a three-year period in Arizona. In ICLARM Conference Proceedings (Philippines). *ICLARM*; *Dept.* of Fisheries.

**Lom**, J. (1986). Cryptobia branchialis Nie from fish gills: ultrastructural evidence of ectocommensal function. *Journal of Fish Diseases*, 3(5), 427-436.

**Lom,** J., & Dyková, I. (2013). Myxozoan genera: definition and notes on taxonomy, lifecycle terminology and pathogenic species. *Folia parasitologica*, 53(1), 1-36.

**Lom**, J., & Dykova, I. (1992). Protozoan Parasites of Fishes. *Elsevier Science Publishers*, Netherlands

**Lom**, J., & Nilsen, F. (2003). Fish microsporidia: fine structural diversity and phylogeny. *International Journal for Parasitology*, 33(2), 107-127.

**Marcotegui**, P. S., Montes, M. M., Barneche, J., Ferrari, W., & Martorelli, S. (2018). Geometric morphometric on a new species of Trichodinidae. A tool to discriminate trichodinid species combined with traditional morphology and molecular analysis. International Journal for Parasitology: *Parasites and Wildlife*,7(2), 228-236.

**Mitchum**, D. L. (1995). Parasites of fishes in Wyoming. Wyoming Game and Fish Dept. **Moratal**, S., Dea-Ayuela, M. A., Cardells, J., Marco-Hirs, N. M., Puigcercós, S., Lizana, V., & López-Ramon, J. (2020). Potential risk of three zoonotic protozoa (Cryptosporidium spp., Giardia duodenalis, and Toxoplasma gondii) transmission from fish consumption. Foods, 9(12), 1913.

**Noga**. E.J. (1996). Fish Disease: Diagnosis and Treatment, Vol. Mosby-Year Book, Inc., *St. Louis, MO*.

**Omeji**, S., Solomon, S. G., & Idoga, E. S. (2011). A comparative study of the common protozoan parasites of Clarias gariepinus from the wild and cultured environments in Benue State, Nigeria. *Journal of Parasitology Research*, 2011.

**Pádua**, S. B., Martins, M. L., Carrijo-Mauad, J. R., Ishikawa, M. M., Jerônimo, G. T., Dias-Neto, J., & Pilarski, F. (2013). First record of Chilodonella hexasticha (Ciliophora: Chilodonellidae) in Brazilian cultured fish: a morphological and pathological assessment. Veterinary Parasitology, 191(1-2), 154-160.

**Paperna**, I. (1980). Parasites, infections and diseases of freshwater fishes in Africa. *CIFA Technical paper*, (7)

**Pavanelli**, G.C.; Eiras, J.C.; Takemoto, R.M. (2008). Doenças de peixes: profilaxia, diagnóstico e tratamento. Maringá: EDUEM, 338p.

**Prasertpol**, J. (1989). Cotton disease of Penaeus monodon. Thai Fisheries Gazette (ISSN01256297). Chantaburi Coastal Aquacult. Devl. Cent. Brackishwater Fish. *Div. Dept Fish.* Thailand. 6 pp.

**Ponpornpisit**, A., Endo, M., & Murata, H. (2000). Experimental infections of a ciliate Tetrahymena pyriformis on ornamental fishes. *Fisheries science*, 66(6), 1026-1031.

**Roberts**, R.J. (2001). Fish Pathology. 3rd (Edn.), *Elsevier Health Sciences*, New York, USA, pp. 492. 13.

**Roberts**, R. J., & Shepherd, C. J. (1997). Handbook of trout and salmon diseases (No. Ed. 3). Fishing News Books, c/o Blackwell Science Ltd.

**Ryan**, U., Hijjawi, N., Feng, Y., & Xiao, L. (2019). Giardia: an under-reported foodborne parasite. *International Journal for Parasitology*, 49(1), 1-11.

**Schäperclaus**, W. (1991). Diseases caused by ciliates. *Fish diseases*. *Amerind Publishing, New Delhi*, 702-725.

**SH**, A., & RA, Y. (2022). Survey of Protozoan Parasites in the Mediterranean Horse Macherel (*Trachurus mediterraneus*) from Zliten Coast, Libya.

**Smit**, N. J., & Davies, A. J. (2005). Intraerythrocytic merogony in Haemogregarina koppiensis (Apicomplexa: Adeleorina: Haemogregarinidae). Folia *Parasitologica*, 52(3), 277.

**Su,** Y., Feng, J., Jiang, J., Guo, Z., Liu, G., & Xu, L. (2014). Trypanosoma epinepheli n. sp.(Kinetoplastida) from a farmed marine fish in China, the brown-marbled grouper (Epinephelus fuscoguttatus). *Parasitology research*, 113(1), 11-18.

**Svobodova**, Z. and Kolarova, J. (2004): A review of the diseases and contaminant related mortalities of tench (Tinca tinca L.). *Journal Veterinary Medicina* – Czech, 49 (1): 19–34.

**Székely**, C., Borzák, R., & Molnár, K. (2018). Description of Henneguya jaczoi sp. n.(Myxosporea, Myxobolidae) from Perca fluviatilis (l.)(Pisces, Percidae) with some remarks on the systematics of Henneguya spp. of European fishes. *Acta Veterinaria Hungarica*, 66(3), 426-443.

Thelohan, P. (1895). Recherches sur les myxosporidies... impr. L. Danel.

**Urawa**, S., Ueki, N., & Karlsbakk, E. (1998). A review of Ichthyobodo infection in marine fishes. Fish *Pathology*, 33(4), 311-320.

**Valladao**, G. M. R., Gallani, S. U., De Padua, S. B., LATERÇA, M., & Pilarski, F. (2014). Trichodina heterodentata (Ciliophora) infestation on Prochilodus lineatus larvae: a host–parasite relationship study. *Parasitology*, 141(5), 662-669.

**Wang**, Q., Yu, Y., Zhang, X., & Xu, Z. (2019). Immune responses of fish to Ichthyophthirius multifiliis (Ich): A model for understanding immunity against protozoan parasites. *Developmental & Comparative Immunology*, 93, 93-102.

**Whitaker**, J. W., Pote, L. M., & Hanson, L. A. (2005). Assay to detect the actinospore and myxospore stages of proliferative gill disease in oligochaetes and pond water. North American Journal of Aquaculture, 67(2), 133-137.

Woo, P. T. (1994). Flagellate parasites of fish. In *Parasitic protozoa* (pp. 1-80). Academic Press.

**Woo**, P. T. K. (1987). *Cryptobia* and cryptobiosis in fishes. *Advances in parasitology*, 26, 199-237.

**Younis**, N. A. E. K. (2012). Studies on some problems of protozoal Infection in freshwater fishes (*Doctoral dissertation*, Cairo University).